Effects of Different Pot Mixtures on spathiphyllum (*Spathiphyllum wallisii* Regel) Growth and Development

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Abstract

The growth of *Spathiphyllum wallisii* Regel plants was evaluated using different pot mixtures (v:v). Plant growth was measured by 11 parameters: leaf area, leaf number, mean shoot length, shoot fresh and dry weight, mean root length, root number, root fresh and dry weight, root volume and number of suckers. Parameters such as leaf area, leaf number, shoot fresh and dry weight, root fresh and dry weight and root length were higher in the media containing only perlite. Mean shoot length was higher in the medium containing 3:1 perlite: sand mixture, 1:3 perlite: sand mixture and only perlite; and root number was higher in the medium containing 3:1 perlite: sand mixture and only perlite. Furthermore, root volume was higher in the medium containing equal perlite: sand mixture and only perlite. The highest number of suckers was obtained in equal leafmold: sand mixture. It is concluded that these differences represent a direct effect on the rooting process and that substrate characteristics are of the utmost importance for the quality of rooted plants.

Keywords: Spathiphyllum wallisii Regel, leaf-mold, perlite, quartz-sand

Introduction

Spathiphyllum is a genus of about 40 species of monocotyledonous flowering plants in the family Araceae, native to tropical regions of the Americas and southeastern Asia. Certain species of Spathiphyllum are commonly known as Spath or Peace Lilies. Several species are popular indoor houseplants. Spathiphyllum cleans indoor air of

many environmental contaminants, including benzene, formaldehyde, and other pollutants. It cleans best at one plant per 10 m³. It lives best in shade and needs little sunlight to thrive. It is watered approximately once a week. The soil is best left moist but only needs watering if the soil is dry. Apart from the function of endogenous physiological and morphological factors which affect root formation in cuttings (Hartman et al., 2002), environmental or exogenous conditions during rooting may play a critical role in the quality of the cutting. One of the most important influential exogenous factors is the physical condition at the basal portion of the cutting, e.g., use of various rooting media (Altman, Freudenberg, 1983). It is generally known that most perennial ornamental plants are propagated by vegetative means using cuttings, layering, grafting, budding, etc. The rate of success in harvesting rooted cuttings depends on physiological age, time of rooting, environmental conditions, i.e., light, temperature and humidity and making use of plant growth regulators (Mamba, Wahome, 2010). Combinations of various media have become especially popular in cutting production of ornamentals (Altman, Freudenberg, 1983). However, considerable differences between the qualities of cuttings grown on various media combinations are evident, depending on the plant species and on the specific environmental conditions of the nursery. Although, effects of different pot mixtures on plant growth and development have been previously investigated (Douglas et al., 2000; Nowak, Strojny, 2003; Samartzidis et al., 2005). Potting media as well as nutritional requirements are the most important factors affecting growth of ornamental plants (El-Naggar, El-Nasharty, 2009). The type of rooting media and their characteristics are of utmost importance for the quality of rooted cuttings (Khayyat et al., 2007). commonly used media for encouraging rooting in cuttings include peat moss, coir, rockwool, vermiculite, perlite, sand, shredded bark, garden soil, leaf-mold, compost, etc. Choice of the used medium components depends on availability of materials, size and type of container, method of watering etc. However, the medium should be free of infectious pathogens, weeds, pests, nematodes, have good water holding capacity and good drainage. Soil-less media have become very popular among propagators because of their consistency, excellent aeration, reproducibility and low bulk density, which reduce shipping and handling costs of the medium itself and of the produced plants (Mamba, Wahome, 2010). In the present

investigation, the effects of different pot mixtures on rooting characteristics of *Spathiphyllum wallisii* plants are studied in greenhouse condition.

Materials and Methods

Media

Thirteen pot mixtures were used for this experiment. The compositions of these media, expressed in volume ratios, were as follows:

L100: Only leaf-mold

S100: Only quartz-sand

P100: Only perlite

L50S50: leaf-mold/quartz-sand (1:1)

L25S75: leaf-mold/quartz-sand (1:3)

L75S25: leaf-mold/quartz-sand (3:1)

P50S50: perlite/quartz-sand (1:1)

P25 S75: perlite/quartz-sand (3:1)

L25P75: leaf-mold/peat moss (1:3)

L50P50: leaf-mold/perlite (1:1)

L75P25: leaf-mold/perlite (3:1)

S25P75: quartz-sand/perlite (1:3)

P33S33L33: perlite/quartz-sand/ leaf-mold (1:1:1)

Rooting condition

Sub-terminal stem cuttings of *Spathiphyllum wallisii* were prepared in mid-October 2010. The stem cuttings were equal in length and each had 4 leaves. After planting, all the cuttings were placed in a greenhouse controlled environment with 16°C night temperature. During autumn, light intensity was reduced to 25 to 30 klx by shading the roof. Plants were "hand watered" during the experiment.

Data recording and analysis

The water-holding capacity and the air space of the substrates were calculated by the method of Verdonck and Gabriels (1992) (Table 1). Root and shoot fresh and dry

weights and leaf area were measured using an Analytical single-pan balance and a leaf area meter (Delta-T Devices Ltd., Burwell, Cambridge, England), respectively. In addition, root and shoot lengths and also root and leaf number were measured at the end of experiment (end of June 2010). Experiments were conducted in a Completely Randomized Design (CRD) with 13 treatments, 4 replications .Analysis of data was carried out using SAS software and Means were compared using LSD at the 5% level.

Table 1. The water-holding capacity and the porosity of the substrates used.

| | Total porosity | Water holding capacity | | |
|-------------|----------------|------------------------|--|--|
| Medium | (%) | (%) | | |
| Quartz-sand | 96 | 36 | | |
| Perlite | 76.7 | 59-68.9 | | |
| Leaf-mold | 91 | 68 | | |

Results

There were significant differences between substrates with regard to the quality of produced roots and developed shoots. Greater leaf area, leaf number, shoot fresh and dry weight; root fresh and dry weights and root length were observed in the media containing only perlite and significant differences were observed between this medium and the other pot mixtures (Figures 1 and 2, Tables 2 and 3). Mean shoot length was higher in each separate media containing P75S25 mixture, P25S75 mixture and P100. Root number was higher in the medium containing P75S25 mixture and P100 (Tables 2 and 3). The media containing only perlite yielded the best results with regard to parameters such as leaf area, leaf number, shoot fresh and dry weight; root fresh and dry weights and root length and root volume (Tables 2 and 3). Greater leaf area was observed in P100and was significantly more than the other media (Table 2). The highest leaf number was observed in P100 mixture. However, no significant differences were observed between P75S25 and P25S75 mixtures (Table 2). Root volume was higher in separate media containing P50S50 mixture and P100 (Table 3). The highest

number of suckers was obtained in L50S50 mixture. The P50S50 mixture yielded almost the same number of suckers as the L50S50 mixture (Table 3).



Figure 1. Long shoots produced on the *Spathiphyllum* cuttings cultured in P100 medium.



Figure 2. Root production on one Spathiphyllum cutting cultured in P100 medium.

Table 2. Effects of growing medium components on leaf area, leaf number, mean shoot length, and shoot fresh and dry weight of *Spathiphyllum*.

| Treatment | Leaf area (cm²) | Number of leaves/plant | Shoot length (cm) | Fresh weight of shoots (g/plant) | Dry weight of shoots (g/plant) |
|-----------|--------------------|------------------------------|----------------------|----------------------------------|--------------------------------|
| L100 | 19.90f* | 2.50f | 5.50cd | 0.70f | 0.05d |
| S100 | 173.11cd | 10.50bc | 11.00b | 5.73de | 0.66cd |
| P100 | 478.88a | 14.75a | 19.25a | 17.35a | 2.43a |
| L50S50 | 124.76def | 7.25def | 7.75bcd | 6.11cd | 0.84c |
| L25S75 | 88.43def | 7.00def | 8.00bcd | 3.08def | 0.33cd |
| L75S25 | 29.98f | 3.00f | 4.00d | 1.10ef | 0.12d |
| P50S50 | 144.46def | 8.25bcd | 10.00bc | 6.10cde | 0.73cd |
| P25S75 | 291.98b | 11.50ab | 16.75a | 13.40ab | 1.83ab |
| P75S25 | 250.85bc | 11.50ab | 17.00a | 11.07bc | 1.62b |
| P50L50 | 42.78ef | 4.00ef | 8.00bcd | 2.05def | 0.22cd |
| P25L75 | 104.21def | 5.750def | 10.00bc | 4.64def | 0.42cd |
| P75L25 | 102.11def | 5.00def | 7.50bcd | 3.62def | 0.38cd |
| P33S33L33 | 94.55def | 5.00def | 6.50bcd | 3.62def | 0.36cd |

^{*}In each column, means followed by same letter(s) are not significantly different using LSD test at 5% level.

Table 3. Effects of growing medium components on root length, root number, root fresh and dry weight, root volume and number of suckers of *Spathiphyllum*.

| Treatment | Number of roots/plant | Root length (cm) | Fresh weight of roots (g/plant) | Dry weight of roots (g/plant) | Root volume (cm³) | Number of suckers/plant |
|-----------|-----------------------------|---------------------|---------------------------------|-------------------------------------|-------------------------|-------------------------------|
| L100 | 8.50de* | 7.37d-g | 6.07d | 0.78c | 15.50bc | 1.00c |
| S100 | 17.25cde | 10.50de | 13.18bcd | 2.32bc | 22.50ab | 2.25bc |
| P100 | 45.00a | 25.00a | 28.17a | 5.35a | 35.25a | 1.75c |
| L50S50 | 27.00bc | 12.75c | 23.35abc | 4.13ab | 21.25ab | 5.50a |
| L25S75 | 6.75e | 4.87g | 4.44d | 0.94c | 6.25c | 1.00c |
| L75S25 | 5.75e | 5.25fg | 2.12d | 0.29c | 5.50c | 0.75c |
| P50S50 | 36.50ab | 13.75c | 13.21bcd | 2.79abc | 21.75a | 4.25ab |
| P25S75 | 33.00ab | 19.75b | 24.55ab | 4.37ab | 22.50ab | 3.25abc |
| P75S25 | 44.00a | 19.62b | 21.64abc | 4.34ab | 21.75ab | 1.50c |
| P50L50 | 7.00de | 6.37efg | 3.49d | 0.77c | 4.25c | 2.50bc |
| P25L75 | 27.00bc | 10.75cd | 14.44bcd | 2.07bc | 11.00bc | 3.00abc |
| P75L25 | 19.50cd | 12.00cd | 10.65cd | 1.86bc | 9.25bc | 1.50c |
| P33S33L33 | 16.00cde | 10.25c-f | 10.70cd | 1.99bc | 6.25c | 2.25bc |

^{*}In each column, means followed by the same letter(s) are not significantly different using LSD test at 5% level.

Discussion

It is confirmed that different media have their own efficiencies. Differences in plant's performance on numerous rooting media can be attributed to a direct effect of the substrate on the basal portion of the cutting. The large differences in the quality of the produced root system and shoot characteristics do indeed indicate the importance of direct effects of the media.

Improved root formation and growth on P100 and P75S25 mixtures might be due to the better aeration and drainage condition and water maintenance capabilities of these substrates compared to other media (Eleni et al., 2001; Mamba, Wahome, 2010; Noguera et al., 2000) which are considered as critical for the first phase of the root

initiation. The presence of the leaves on the cuttings may cause earlier growth of the root system, but other environmental factors can also be involved. Thus, while new leaf development on P100 mixture is largely concurrent with the superior root development on these media, L100 has less deleterious effects on leaf growth.

On the other hand, sand mixtures allowed moderate leaf development, although root growth was not noticeable. In the media containing leaf-mold, low leaf development occurred; also, root growth was low. Since these phenomena cannot be explained solely by differences in the water/air relationship of the various rooting media, other factors are probably involved. Mechanical impedance and reduced porosity are such factors which may restrict root formation (Khayyat et al., 2007).

The number of leaves produced per cutting is determined by the type of cutting used, utilized plant growth regulators, temperature, and dry matter content of the cuttings before planting in the medium and the health status of the plant. Since all cuttings used in this investigation were uniform, the highest number of leaves per cutting observed in *Spathiphyllum wallisii* plants rooted in the mixture of perlite and sand could be attributed to other medium characteristics like porosity and water holding capacity. In conclusion, perlite was found to be superior in the propagation of *Spathiphyllum wallisii* plants as compared to the other medium components when most of the root and shoots parameters were evaluated. Only perlite and the mixture of perlite and sand, are therefore recommended for use in commercial propagation of *Spathiphyllum wallisii* plants under suitable environmental conditions.

References

- Altman, A., Freudenberg, D. (1983) Quality of *Pelargonium graveolens* cutting as affected by the rooting medium. Scientia Horticulturae 19, 379-385.
- Douglas, M.H., Smallfield, B.M., Parmenter, G.A., Burton, L.C., Heaney, A.J. (2000)

 Effect of growing media on the production of ginseng (*Panax ginseng*) in Central Otago, New Zealand. New Zealand Journal of Crop and Horticultural Science 28, 195-207.
- Eleni, M., Sabri, K., Dimitra, Z. (2001) Effect of growing media on the production and quality of two rose varieties. Acta Horticulturae 548, 79-83.

- El-Naggar, A.H., El-Nasharty, A.B. (2009) Effects of growing media and mineral fertilisation on growth, flowering, bulb productivity and chemical constituents of *Hippeastrum vittatum*, Herb. American-Eurasian Journal of Agricultural and Environmental Science 6, 360-371.
- Hartman, H.T., Kester, D.E., Davies, F.T., Geneve, R.L. (2002) Plant Propagation,
 Principles and Practices. Pearson Education, Inc., Upper Saddle River, NJ, USA
 880 p.
- Khayyat, M., Nazari, F., Salehi, H. (2007) Effects of different pot mixtures on Pothos (*Epipremnun aureum* Lindl. and Andre 'Golden Pothos') growth and development. American-Eurasian Journal of Agricultural and Environmental Science 2, 341-348.
- Mamba, B., Wahome, P.K. (2010) Propagation of geranium (*Perlagonium hortorum*) using different rooting medium components. American-Eurasian Journal of Agricultural, Environmental Science 7, 497-500.
- Nicolosi, R.T., Fretz, T.A. (1980) Evaluation of root growth in varying medium densities and through dissimilar soil surfaces. HortScience15, 642-644.
- Noguera, P., Abad, M., Noguera, V., Puchades, R., Maquieira, A. (2000) Coconut coir waste, a new and viable ecologically friendly peat substitute. Acta Horticulturae 517, 279-286.
- Nowak, J.S., Strojny, Z. (2003) Effect of different container media on the growth of gerbera. Acta Horticulturae 608, 59-63.
- Samartzidis, C., Awada, T., Maloupa, E., Radoglou, K., Constantinidou, I.A.H. (2005)

 Rose productivity and physiological responses to different substrates for soil-less culture. Scientia Horticulturae106, 203-212.
- Verdonck, O., Gabriels, R. (1992) I. Reference method for the determination of physical properties of plant substrates. II. Reference method for the determination of chemical properties of plant substrates. Acta Horticulturae, 302: 169-179.